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### Mycorrhizal symbioses and plant interactions

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### Keywords

Plants interaction; Symbiotic microorganisms; Microbial biodiversity; Rhizosphere

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### REVIEW ARTICLE Mycorrhizal Symbioses and Plant Interactions

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#### Abstract

The growing interest in mycorrhizal fungi in agriculture is related to their symbiotic relationships with cultivated plants. Thanks to functional genomics approaches, mycorrhizae and symbioses with host plants have emerged for their features. Besides improving nutritional supply, plant-fungal interactions increase plants' tolerance to abiotic stresses such as drought, salinity and cold, as well as their resistance to diseases. Recent studies have investigated the interactions between plants and mycorrhizae, however the mechanisms often remain unclear. Indeed, plants in the field are affected by various stresses and results often appear contradictory. This review is aimed at presenting the most relevant studies in this field in order to highlight the possible benefits of mycorrhizal interactions and their application in agriculture.

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#### 1. Introduction

owards the end of the 19th century, symbiosis was understood as a long-lasting association established by different species [1]. Symbiosis encompasses a broad spectrum of relationships involving benefits for both partners, antagonistic relationships and relationships from which one organism benefits [2,3]. The type of association often depends on the type of environment and the developmental stage of the partners. The basis of symbiosis is undoubtedly the possibility of an organism acquiring new metabolic capabilities from the relationship established with its new partner. Symbioses represent a solid evolutionary and innovative significance in the development of new organs, tissues and cells [4]. In the interaction that can take place between nitrogen-fixing microorganisms and leguminous plants, a series of events occur that ensure the relationship between the two organisms: the presence of signal molecules, a signal uptake system, the activation of transcription factors and target genes [5]. Plants has an underdeveloped root system, which makes arbuscular mycorrhizae a crucial factor in plant evolution and territory conquest [6]. The ability of symbiont micro-organisms to enhance the mineral uptake of plants, especially in nutrient-poor environments, certainly favoured symbiotic relationships [7]. Unlike arbuscular mycorrhizae, ectomycorrhizae seem to have appeared much later. These fungi and their symbiotic relationships have adapted to environments rich in organic matter, so it can be outlined that the type of nutrient exchange certainly depends on the type of mycorrhiza and reflects the different ecological specialisations of these associations [8]. Mycorrhizae are not obligatory relationships with plants, as the latter can also grow without the fungus, especially in nutrient- and water-rich soils, such as those fertilised by agriculture. Nervetheless, the mycorrhizal arbuscules can develop symbiotic relationships with their host plants that determine their dependence. Mycorrhizae play a crucial role in the establishment and stability of plant communities [9-11]. The role of mycorrhizal symbiosis in plant communities and ecosystems has received more attention in recent research. In fact, it has become increasingly important to study the extraradical mycelial phase of the symbiosis and to use realistic substrates in answering pertinent ecological questions [12,13]. Many authors have highlighted the multifunctional nature of mycorrhizal effects [14,15], including

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interactions with bacteria [16], weathering of minerals [17-19], carbon cycling [20], effects on plant communities [21], tripartite syntheses with mycoheterotrophs [22,23], and mediation of plant responses to stress [24]. Through symbiosis, the plant improves its mineral nutrition, having a positive effect on its growth, in exchange for carbon compounds that are given to the fungus [25,26]. The success of mycorrhizal fungi, in time and space, is mainly related to the nutritional benefits they confer on their host plants: through the extraradical mycelium they absorb phosphate and other macronutrients, as well as trace elements and water from the soil, and release them to the plant [27]. Specifically, nutrients are exchanged through the symbiotic interface between the plant and the fungus, which is created within the roots [25,27-30]. Moreover, using the symbiosis, the plant becomes more resistant to biotic stresses [31] and increases tolerance to abiotic stresses, such as lack of water or the presence of pollutants [28,29]. In recent years, it has also been highlighted that the effects of symbiosis on plant response may be different and may be related to the genotypes of the two symbiotic organisms, environmental conditions as well as the availability of resources [32].

The aim of this review is to highlight the mechanisms and effects that underlie the symbiosis between plants and mycorrhizae, in order to gain a better understanding of these fundamental relationships in plant life and for soil biodiversity and fertility. There is still little understanding of the detailed functioning and regulation of these mycorrhizosphere processes, but recent progress has been reviewed, as well as the potential benefits of an improved understanding of mycorrhizosphere interactions.

#### 2. The various mycorrhizal symbioses

Generally, mycorrhizal symbioses can occur for several reasons: anatomical characteristics, the type of nutrients exchanged, and the taxonomic position of the organisms involved [33]. In order to determine the type of mycorrhizal symbiosis, it is generally observed how the symbiont fungus affects the host plant cells. By contrast, ectomycorrhizal fungi remain outside plant cells, while endomycorrhizal fungi colonize root cells. In the ectomycorrhizal symbiosis, the fungus produces a layer of hyphae called mycoclena around the root, from which organized hyphae branch off to explore the soil and transport water and nutrients. At the same time, a hyphal network develops on the root, which surrounds the plant cells without penetrating them (Hartig's network) [34,35]. A similar process occurs in endomycorrhiza, where extraradical hyphae explore the soil and absorb nutrients, however the process is not comparable to ectomycorrhiza [36]. Arbuscular mycorrhizae take their name from the arbuscule, a structure generated by branching the hypha within cortical cells. There are also other types of symbiosis, in which fungi generate hyphal galls, as in the case in orchids and ericoid mycorrhizae (Table 1) [37]. Ectoendomycorrhizae present characters intermediate between ectomycorrhizae and endomycorrhizae and are specialized in a limited number of plant species, on which they generally develop a mycoclena or Hartig reticulum associated with the root penetration of the fungus [38,39]. Ectomycorrhizae are found in deciduous and coniferous woodland environments in temperate and cold climates. The soil of temperate forests generally has a thick layer of organic material from leaf litter, rich in compounds that are difficult to degrade [40]. In these soils, nitrogen is in organic form, easily absorbed by ectomycorrhizal fungi and transferred to the host plant. In contrast, arbuscular mycorrhizae are present in temperate and warm climates. In these areas, microbial activity quickly degrades organic matter, and phosphorous is the limiting element for plant growth [41,42]. Therefore, arbuscular mycorrhizal fungi mainly specialize in the transfer of phosphates in inorganic form. Three different forms of mycorrhiza can be identified in the Ericaceae. These plants are characterized by having excellent roots, and generally, the epidermis is the cell layer colonized by the fungi. Erycoid mycorrhizae are mainly found in tundra and heathland environments both in Mediterranean and cold regions. In heaths strongly characterized by acidic soils, mineralization processes are mainly slowed down, and the availability of heavy metals can be high [43]. Under these conditions, the plants' resistance is significantly increased by their association with symbiotic fungi, which enable the breakdown of complex molecules and transfer of nutrients in organic form to the plants [44,45]. Plants in the soil exhibit mycorrhiza consistently

Table 1. List of certain ectomycorrhizal fungi [37].

Ectomycorrhizal symbionts	Family
Suillus luteus (L.) Roussel	Suillaceae
Suillus granulatus (L.) Roussel	Suillaceae
Boletus edulis Bull.	Boletaceae
Amanita muscaria (L.) Lam.	Amanitaceae
Clitopilus prunulus (Scop.) P. Kumm.	Entolomataceae
Lactarius delicious (L.) Gray	Russuluceae
Craterellus lutescens (Pers.) Fr.	Cantharellaceae

regardless of the ecological area in which they grow, whether they are epiphytes or terrestrials [46]. Orchids are characterized by a complex life cycle. Plants in the heterotrophic phase cannot photosynthesize since photosynthetic pigments are absent and rely on mycorrhizal fungi for carbon acquisition. For this reason, mycorrhizae represent a standard and, in many cases, a fundamental element in the life of terrestrial species. Their first appearance coincided with the evolution of host plants [47,48]. In addition to mycorrhizal fungi's crucial nutritional role, they defend the host plant from numerous abiotic and biotic stresses [49].

### 3. Determining mechanisms in biological biodiversity

Mycorrhizal biodiversity at the root level closely reflects the specificity of the particular fungus-plant association [50]. In the different mycorrhizal types, very different situations are observed regarding the diversity of fungal symbionts. In ectomycorrhizae, an impressive richness of fungal symbionts species is observed, requiring the host plant to complete its life cycle and produce a fruiting body [51,52]. Measuring the number of fruiting bodies produced by each species within the plant community has long been used to assess the diversity and relative abundance of different ectomycorrhizal species. The first study on symbionts showed a massive discrepancy in the production of fruiting bodies, as species frequently found as symbionts produced significantly few fruiting bodies [53,54]. In contrast, other species found as fruiting bodies were rarely associated with root apices [55]. Studies have thus shown that a small number of ectomycorrhizal fungal species colonize up to 70% of root apices, while many species appear more rarely [56]. The composition of these fungal communities varies depending on numerous biotic and abiotic factors [57]. The plant benefits obtained from symbioses with ectomycorrhizal fungi are attributable to access to otherwise inaccessible nutrients [58,59]. They produce a wide range of extracellular enzymes capable of decomposing complex organic molecules. The fungus can recover nitrogen and phosphorous in an organic form that can be partly given to the plant in exchange for sugars [60]. Since each fungus strain produces its spectrum of enzymes with different biochemical characteristics, the advantage of associating with a variety of fungal symbionts is quite evident for a plant growing in nutrient-poor ecosystems rich in organic compounds of a varied and complex nature [61-63]. Compared to ectomycorrhizae, where fungal species are numerous and belong to different taxa, arbuscular mycorrhizae

have around a hundred species of symbionts. Several recent studies have demonstrated that plants and arbuscular mycorrhizae' biodiversity is closely related [64]. Microcosm studies have shown that the species composition of the plant community correlates with the type of fungi present. The plants most favoured by establishing an affluent fungal population depend on mycorrhiza for mineral nutrition and growth (Fig. 1) [65]. The increase in diversity observed in plant communities following the increase in fungal species primarily reflects an increased presence of these mycorrhiza-dependent species. Concerning the diversity of fungal symbionts, very different conclusions have been reached by identifying mycelia isolated from mycorrhized roots or by molecular analysis [66]. Regarding the diversity of fungal symbionts, very different conclusions have been reached by identifying mycelia isolated from mycorrhized roots or by molecular analysis [67]. In the heterotrophic phase of the life cycle, the plant acquires organic carbon compounds by exploiting its symbionts' strong degradative ability to degrade organic matter in the environment [68]. These methods have been used to identify the mechanism of acquiring organic carbon compounds. In mycorrhizal symbiosis, filamentous structures are composed of symbiont fungi. On the one hand, they interact with plant roots and explore the soil extensively, even over considerable distances. This unique cellular organization makes mycorrhizal symbiosis a fascinating phenomenon [69]. Alternatively, in this exploration phase in the soil, fungal hyphae can make contact with the roots of other potential hosts, forming hyphal connections between the roots of distinct plant individuals [70]. Depending on the specificity of the plant-fungus interaction, these hyphal connections may involve individuals of the same or different species. Different types of mycorrhizal fungi transport different nutrients: arbuscular mycorrhizal fungi transport mainly phosphorous and inorganic nitrogen. As opposed to this, ectomycorrhizal fungi transfer organic carbon compounds [30,71]. Studies concerning these interconnections have revealed

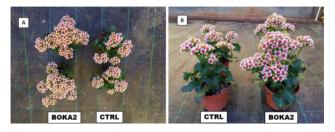


Fig. 1. Improving plant nutrition through mycorrhizal symbioses can increase flowering in Kalanchoe blossfeldiana. Boka2 (TREATED) vs CTRL (CONTROL).

some consequences of the distribution of organic carbon resources [72], for example, in transferring sugars between photosynthetic adult plants and young seedlings, which would otherwise die. In *Crocus sativus* plants, the different types of mycorrhizae (ectomycorrhizae and endomycorrhizae) have a different effect on the vegetative and root development of the plants (Fig. 2). Most heterotrophic plants have evolved this sophisticated exchange mechanism through the fungi's network of hyphal interconnections to obtain the needed sugars. Moreover, several species have evolved a similar mechanism, indicating evolutionary convergence [73].

# 4. The different interactions between plant and symbiont fungus

Despite the genetic and functional multiplicity, there is a unifying factor in that the host plant must accommodate the fungus in the root, giving rise to micromorphogenesis events that can only be appreciated microscopically. It can be observed that the modification of the root is the result of coordinated colonization processes.

- 1) development of the fungus in the rhizosphere and the exchange of signals with the plant;
- 2) contact with the root surface and the development of adhesion structures (Fig. 3)
- 3) development of fungal structures associated with the root surface and intraradical.

The mycorrhizal symbiosis phase represents a stage in a very complex life cycle comprising.

- a saprotrophic phase where mycelia originating from spores of the sexual origin or vegetative propagules grow even in the absence of the host plant and proliferate in the soil;
- ii) a symbiotic association phase;



Fig. 2. Effect of different types of mycorrhiza Laccaria bicolor (LB) ectomycorrhiza and Glomus deserticola (GD) endomycorrhiza, on root and vegetative growth of Crocus sativus.

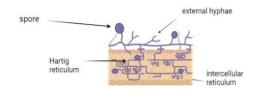


Fig. 3. Process of colonisation of the root by mycorrhizae with description of their structures.

iii) a reproductive phase culminating in the production of an ascocarp or basidiocarp.

In the molecular interaction between the two partners, the fungus contacts the root surface, and the hyphae increase in diameter and branch out, cleaving the median lamella between the epidermal cells and insinuating themselves, forming the first elements of the Hartig reticulum [74]. As a result of this morphological organization, functions are compartmentalized: extraradical hyphae function as nutrient uptake structures; the mantle serves as a reserve structure for nutrients; the Hartig reticulum serves as a site for exchange between both hosts due to its increased contact surface. In some fungal genotypes, defence reactions can also occur, with an accumulation of callose. It is clear, therefore, that there are essential processes that change the morphology of the root and the shape of the epidermal cells in the type of growth regulated by genes and signal exchanges in the rhizosphere [75]. The genetic determinants that control the process of mycorrhizal symbiosis in plants are beginning to be known through the study of mutants, which have enabled the identification of a good number of genes that perform similar functions in different plants have highly conserved sequences. In controlling the colonization process, the secondary metabolism of host plants may be of great importance [76]. It has long been known that flavonoids released from roots can enhance the growth process of hyphae from germinated spores, but the role of these compounds is still unclear. Recently, however, it has been shown that the release of metabolites into the soil stimulates hyphal branching, simultaneously the respiratory process, and other related oxidative metabolisms [77].

# 5. Molecular mechanisms determining the establishment of symbioses

Knowledge of the molecular mechanisms that enable the establishment of symbioses has leapt forward thanks to the development of molecular biology technologies [78]. Through them, it has been possible to identify genes of interest, analyze their functions and assess gene expression under different environmental conditions [79]. Various scientific tests have shown that mycorrhized plants grow better than non-mycorrhized ones [30]. This is generally related to the improved uptake of nutrients mediated by the symbiont fungi, which is particularly important for poorly mobile ions such as phosphate and ammonium [80]. These ions are known to be growth-limiting for plants, as they become deficient in the rhizosphere, where a depletion zone forms, following uptake by the root hairs [81]. The hyphae of the symbiont fungi can efficiently counterbalance cuttings depletion area, as nutrients through the fungal hyphae pass to the root cells acting as sinks faster than they diffuse into the soil. This faster translocation activity explains the increased uptake rate of mycorrhized roots [82]. The improved vegetative development of mycorrhized plants is primarily due to the improved availability of phosphorus (Fig. 4). The symbiont fungi can take up the free phosphorus in the soil beyond the depletion zone and transfer it to the plant [83]. Mycorrhizal fungi can solubilize forms of phosphorous that the plant cannot solubilize, such as calcium and aluminium phosphates, thanks to the controlled secretion of extracellular phosphatases [84]. A gene coding for a high-affinity transporter for phosphate, especially active in outer hyphae, is present in Glomus spp. [85]. Even in the same ecosystem, mycorrhizal fungi can explore different niches and exploit mineral element sources differently. It is especially true for nitrogen: significant changes are influenced by the concentration, form, and availability of plants, all of which correlate with soil microorganism degradation cycles (Fig. 5) [86]. At low temperatures and acid pH, nitrogen mineralization is so slow that organic sources become essential. In addition to having high-affinity ammonium transporters similar to yeast ammonium permeases MEP2, ectomycorrhizal fungi possess very active ammonium uptake agents [87]. Activated transporters occur during nitrogen



Fig. 4. Increased plant height and leaf growth in Plectranthus amboinicus plants treated with Glomus intraradices under phosphorus deficient conditions. The growth is evident in treated plants (AM), compared to control plants (CTRL).

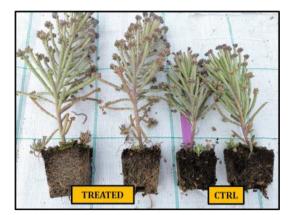


Fig. 5. Stimulating effect of Glomus mossae on Kalanchoe daigremontiana plants in the presence of nitrogen. Increased vegetative and root growth and number of new shoots in the treated thesis (TREATED) compared to the control (CTRL) under conditions of reduced nitrogen content in the growing medium.

deficiency, are repressed by glutamine, and are active during the symbiotic phase. It is still being determined how glutamine synthetase is transferred to plants [88]. Several studies have indicated glutamine synthetase to be highly expressed in some symbionts during nitrogen deficiency and fruiting body formation [89]. A gene expression study has shown that gene determinants are only activated when the fungus emerges on a sugar-poor medium and is low in glucose and ammonia. Therefore, assimilation pathways differ significantly depending on how closely the plant is associated with the host [90]. The molecular approach was used to identify nitrate reductase genes expressed by mycorrhizal maize roots and distinguish between symbiont genes and host genes [91].

#### 6. Synergistic activity of soil microorganisms

Several analyses have recently been conducted on the relationship between soil microorganisms and symbiotic fungi, demonstrating that they can significantly impact plant health because of the synergy between various populations in the rhizosphere [92]. In order to explain these interactions, many mechanisms have been proposed, including phytohormone production, enzyme release, vitamin release, and growth stimulation. Mycorrhizae and rhizobacteria may be able to form biofilms on the surface of hyphae, preventing their leaching and, ultimately, their loss of the inoculum. In light of this, increased bacteria numbers and concentrations locally may explain the beneficial effects [93]. Recent studies have shown that certain Pseudomonas strains can adhere to fungal hyphae due to the presence of specific molecules such as mucopolysaccharides [94]. Some mycorrhizae also

harbour several bacteria in their cytoplasm, which, though unculturable with small genome size, are transmitted vertically to future generations. As a result of this symbiosis within symbiosis, the fungal genome is more complex due to the resident genomes [95]. Symbioses between microorganisms are fascinating because of their peculiar cellular organization, which has structures capable of interacting with the roots of host plants and exploring the soil around them. Depending on the specificity of the microorganism-plant interaction, these connections may involve individuals of the same or different species [94]. Depending on the environment, these lattices can move organic and inorganic nutrients from one plant to another (Table 2). According to these relationships in the subsoil, the concept of plant communities as distinct individuals competing with one another is profoundly altered. Rather than acting antagonistically, these individuals symbiosis to provide nutrients as part of their resources [96].

### 7. Importance of exploiting soil interactions between stimulating microorganisms

In response to the adverse effects of plant protection products and synthetic fertilizers on agrosystems, strategies have been developed to promote beneficial soil microorganisms [97]. Consequently, microbial inoculation is of utmost importance because the soil microbiota can play an essential role in plant growth. As a result of microbiota's beneficial effects on plant health under biotic and abiotic stress conditions, biotechnology applications are relevant. In recent years, mycorrhizal growth and health have been highlighted, as well as the nutritional value of edible parts of various crops [98]. Mycorrhizae are not only a simple substitute for chemical fertilizers or pesticides but also can be used in food products of organoleptic and nutraceutical quality despite a lack of comprehensive research on their systemic effects on edible plants. Various vegetables were shown to yield more and be healthier with mixed inoculations of mycorrhizae and growth-promoting bacteria. Similar results were obtained for strawberries and perennials, including olives and vines [99]. Mycorrhizal fungi have been studied on some ornamental plants, especially Camellia japonica, with significant effects on growth and protection [100]. The development of large-scale sequencing approaches has led to the identification and selection of isolates forming symbiotic relationships with plants in different environments or soils subjected to different cultivation techniques, allowing researchers to identify and select symbiotic fungi. Identifying isolates for a specific crop or already adapted to certain conditions and, thus, potentially more efficient depends on data on the composition of mycorrhizal fungi communities [101,102].

# 8. Benefits of introducing mycorrhizal fungi into the soil

The enhancement of P uptake is generally considered to be the most important benefit that mycorrhizae provide to the host plant, and the P status of the plant is often the main controlling factor in the plant-fungus relationship [103]. In some cases, mycorrhizae can increase P uptake and increase the performance efficiency of P nutrition of crops [104]. Increased yield and growth can be associated with this phenomenon [104]. It is common for several nutrients to be absorbed by the host simultaneously by mycorrhizal fungi, though different nutrients may be affected differently [105,106]. The mycorrhizal association has been extensively studied, but there is also evidence that mycorrhizae play a role in the suppression of crop pests and diseases, particularly soil-borne fungal diseases (Table 3) [107,108]. As a result of improved nutrition, plants may appear resistant to pests and diseases [109], although multiple mechanisms of resistance may be at work simultaneously [110]. Exclusion appears to be one of the most important factors here, as it seems to be merely a matter of competing for space [111]. Mycorrhizae have been

Table 2. Nutritional exchanges between different types of mycorrhizae.

Mycorrhizal typology	Symbiont > host nutritional interaction	Host > symbiont nutritional interaction
Arbuscular mycorrhizae	Mineral nutrients (P)	Carbohydrates
Ectomycorrhizae	Mineral nutrients (N)	Carbohydrates
Hericoids	Mineral nutrients (N)	Carbohydrates
Arbutoids	Mineral nutrients	Carbohydrates
Monotropoids	Carbohydrates and mineral nutrients	
Orchid mycorrhizae	Carbohydrates and mineral nutrients	_
-	during the young phase; minerals	
	during the adult phase	

Table 3. Mycorrhizae control some soil fungal diseases.

Pathogen	Disease	Crop
Sclerotium cepivorum	White rot	Onions (Allium cepa)
Fusarium oxysporum	Fusarium root rot	Asparagus (Asparagus officinalis)
Verticillium dahlie	Verticillium wilt	Tomatoes (Lycopersicon esculentum)
Helicobasidium mompa	Violet root rot	Asparagus
Rhizoctonia solani	Root and stem rots	Mung bean (Vigna radiate)
Aphanomyces euteiches	Root rot	Pea (Pisium sativum)

shown to increase the host plant's tolerance to water stresses [112], including those caused by high salinity [106,113,114]. Several mechanisms have been proposed to explain why water is extracted from smaller pores [112]: increased hydraulic conductivity of the roots, improved stomatal regulation, osmotic regulation of the host and better contact with soil particles due to the binding effect of hyphae). Many studies have focused on how mycorrhizae directly influence the growth of the host plant. They also have a direct effect on soil structure, which is particularly relevant in agricultural environments, in which cultivation, traffic and low levels of organic matter tend to damage soil structure. According to Jakobsen and Rosendahl [115], host plants can transfer up to 20% of fixed carbon to their fungal partners and mycorrhizae can produce significant biomass in agricultural soils [116].

#### 9. Conclusions

The molecular mechanisms underlying the exchange of nutrients between two symbiont organisms are rapidly emerging and a nutritional advantage for the host plant has been established. to the development of large-scale Thanks sequencing approaches, several researches have been focused on the characterisation of mycorrhizae in different environments or in subsoils characterized by different cultivation techniques. In recent years, mycorrhizal symbiotic interactions have been studied in greater depth, and it appears that nutritional exchanges and physiological responses of plants may depend on the environment and the partners involved in the symbiotic relationship, at least in mycorrhizal symbioses. In addition, mycorrhizal fungi could regulate the process of signal transfer between plants above ground, as well as the transfer of signalling molecules between plants. In order to optimize the use of these fungi in application programmes, all of these aspects will have to be investigated in the future. Many of these studies have also been carried out in roots sampled in natural soils, making it possible to identify isolates that form symbioses with the plants under consideration and making it

possible to observe that not all fungi associated with roots form a functional symbiosis. Despite much research, on the whole, the beneficial properties of mycorrhizal symbioses are still not widely exploited in sustainable agriculture. This is because many factors must be considered in order to achieve successful application. In order to be used for agricultural purposes, mycorrhizae must also be evaluated nutritionally and physiologically in fields with different soil microbiota communities, with which both symbiont fungi and plant roots interact, as well as multiple stress factors affecting plants. Although we are beginning to understand the association of microorganisms and plants, there are significant gaps, including the role of species diversity of bacteria and fungi in producing plant benefits, as well as how agronomic practices can affect the ecology and function and development of beneficial microorganisms in the soil. It is crucial to improve our understanding of the physiology and function of mycorrhiza, as well as their interactions with crops and environmental conditions, to achieve long-term agricultural stability and productivity.

The study of the specific plant-fungus-environment combination is an important prerequisite, as well as the implementation of less impactful agricultural practices so that the biodiversity of this group of fungi is maintained in the soil.

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#### References

- [1] D. Mbodj, B. Effa-Effa, A. Kane, B. Manneh, P. Gantet, L. Laplaze, A.G. Diedhiou, A. Grondin, Arbuscular mycorrhizal symbiosis in rice: establishment, environmental control and impact on plant growth and resistance to abiotic stresses, Rhizosphere 8 (2018) 12–26.
- [2] E.C.H. Chen, É. Morin, D. Beaudet, J. Noel, G. Yildirir, S. Ndikumana, P. Charron, C. St-Onge, J. Giorgi, M. Krüger, High intraspecific genome diversity in the

model arbuscular mycorrhizal symbiont *Rhizophagus irregularis*, New Phytol. 220 (2018) 1161–1171.

- [3] Y.N. Zou, Q.S. Wu, K. Kuca, Unravelling the role of arbuscular mycorrhizal fungi in mitigating the oxidative burst of plants under drought stress, Plant Biol. 23 (2021) 50-57.
- [4] J. Choi, W. Summers, U. Paszkowski, Mechanisms underlying establishment of arbuscular mycorrhizal symbioses, Annu. Rev. Phytopathol. 56 (2018) 135–160.
- [5] A. Bahadur, A. Batool, F. Nasir, S. Jiang, Q. Minsen, Q. Zhang, J. Pan, Y. Liu, H. Feng, Mechanistic insights into arbuscular mycorrhizal fungi-mediated drought stress tolerance in plants, Int. J. Mol. Sci. 20 (17) (2019) 1–18.
- [6] A. Fusconi, M. Mucciarelli, How important is arbuscular mycorrhizal colonization in wetland and aquatic habitats? Environ. Exp. Bot. 155 (2018) 128–141.
- [7] A. Nanjundappa, D.J. Bagyaraj, A.K. Saxena, M. Kumar, H. Chakdar, Interaction between arbuscular mycorrhizal fungi and *Bacillus* spp. in soil enhancing growth of crop plants, Fungal Biol. Biotechnol. 6 (2019) 1–18.
- [8] S. Primieri, J.C.P. Santos, P.M. Antunes, Nodule-associated bacteria alter the mutualism between arbuscular mycorrhizal fungi and N2 fixing bacteria, Soil Biol. Biochem. 154 (2021) 108–149.
- [9] D.A. Ferreira, T.F. da Silva, V.S. Pylro, J.F. Salles, F.D. Andreote, F. Dini-Andreote, Soil microbial diversity affects the plant-root colonization by arbuscular mycorrhizal fungi, Microb. Ecol. 82 (2021) 100–103.
  [10] R. Roth, U. Paszkowski, Plant carbon nourishment of
- [10] R. Roth, U. Paszkowski, Plant carbon nourishment of arbuscular mycorrhizal fungi, Curr. Opin. Plant Biol. 39 (2017) 50-56.
- [11] L.H. Luginbuehl, G.N. Menard, S. Kurup, H. van Erp, G.V. Radhakrishnan, A. Breakspear, G.E.D. Oldroyd, P.J. Eastmond, Fatty acids in arbuscular mycorrhizal fungi are synthesized by the host plant, Science 356 (2017) 1175–1178.
- [12] Ř.D. Finlay, Mycorrhizal fungi and their multifunctional roles, Mycologist 18 (2004) 91–96.
- [13] I.C. Anderson, J.W.G. Cairney, Ectomycorrhizal fungi: exploring the mycelial frontier, FEMS (Fed. Eur. Microbiol. Soc.) Microbiol. Rev. 31 (2007) 388–406.
- [14] K.K. Newsham, A.H. Fitter, A.R. Watkinson, Multi-functionality and biodiversity in arbuscular mycorrhizas, Trends Ecol. Evol. 10 (1995) 407–411.
- [15] J. Johansson, L. Paul, R.D. Finlay, Microbial interactions in the mycorrhizosphere and their significance for sustainable agriculture, FEMS (Fed. Eur. Microbiol. Soc.) Microbiol. Ecol. 48 (2004) 1–12.
- [16] J. Frey-Klett, J. Garbaye, M. Tarkka, The mycorrhiza helper bacteria revisited, New Phytol. 176 (2007) 22–36.
- [17] R. Landeweert, E. Hofflund, R.D. Finlay, N. van Breemen, Linking plants to rocks: ectomycorrhizal fungi mobilize nutrients from minerals, Trends Ecol. Evol. 16 (2001) 248–254.
- [18] R.D. Finlay, A. Rosling, G.M. Gadd, Integrated Nutrient Cycles in Forest Ecosystems, the Role of Ectomycorrhizal Fungi, Fungi in Biogeochemical Cycles, UK Cambridge University Press. 2006, pp. 28–50.
- [19] H. Wallander, G.M. Gadd, Mineral Dissolution by Ectomycorrhizal Fungi, Fungi in Biogeochemical Cycles, UK Cambridge University Press. 2006, pp. 28–50.
- [20] D. Johnson, J.R. Leake, N. Ostle, P. Ineson, D.J. Read, In situ CO2 pulse-labelling of upland grassland demonstrates that a rapid pathway of carbon flux from arbuscular mycorrhizal mycelia to the soil, New Phytol. 153 (2002) 327–334.
- [21] M.G.A. van der Heijden, J.N. Klironomos, M. Ursic, P. Moutoglis, R. Streitwolf-Engel, T. Boller, A. Wiemken, I.R. Sanders, Mycorrhizal fungal diversity determines plant biodiversity, ecosystem variability and productivity, Nature 396 (1998) 69–72.
- [22] J.R. Leake, Myco-heterotroph/epiparasitic plant interactions with ectomycorrhizal and arbuscular mycorrhizal fungi, Curr. Opin. Plant Biol. 7 (2004) 422–428.

- [23] M.I. Bidartondo, The evolutionary ecology of myco-heterotrophy, New Phytol. 167 (2005) 335–352.
- [24] R.D. Finlay, B.D. Lindahl, A.F.S. Taylor, S. Avery, M. Stratford, P. van West, Responses of Mycorrhizal Fungi to Stress, Stress in Yeasts and Filamentous Fungi, Amsterdam Elsevier. 2008, pp. 201–220.
- [25] S.E. Smith, D.J. Read, Mychorrhizal Symbiosis, Academic Press, Cambridge, UK. 2008, pp. 100–156.
- [26] M.G.A. van der Heijden, F.M. Martin, M. Selosse M., I.R. Sanders, Mychorrhizal ecology and evolution: the past, the present, and the future, New Phytol. 205 (2015) 1406–1423.
- [27] M.K. Rick, E. Nouri, P.E. Courty, D. Reinhardt, Diet of arbuscular mycorrhizal fungi: bread and butter? Trends Plant Sci. 22 (2017) 652-660.
- [28] R. Balestrini, P. Bonfante, Cell wall remodeling in mycorrhizal symbiosis: a way towards biotrophism, Front. Plant Sci. 5 (2014) 1–10.
- [29] R. Balestrini, E. Lumini, Focus on mycorrhizal symbioses, Appl. Soil Ecol. 123 (2018) 299–304.
- [30] M. Chen, M. Arato, L. Borghi, E. Nouri, D. Reinhardt, Beneficial services of abruscular mycorrhizal fungi. From ecology to application, Front. Plant Sci. 9 (2018) 1–14.
- [31] M.J. Pozo, A. Verhage, J. Garcia-Andrade, J.M. Garcia, C. Azcon-Aguilar, Priming plant defence against pathogens by arbuscular mycorrhizal fungi, Processes and Ecological Impact (2009) 123–135.
- [32] A. Berruti, A. Desirò, S. Visentin, O. Zecca, P. Bonfante, ITS fungal barcoding primers versus 18S AMF-specific primers reveal similar AMF-based diversity patterns in roots and soils of three mountain vineyards, Environmental Microbiology Reports 9 (2017) 658–667.
- [33] C. Liu, S. Ravnskov, F. Liu, G.H. Rubæk, M.N. Andersen, Arbuscular mycorrhizal fungi alleviate abiotic stresses in potato plants caused by low phosphorus and deficit irrigation/partial root-zone drying, J. Agric. Sci. 156 (2018) 46–58.
- [34] G. Winkelmann, A Search for Glomuferrin: a potential siderophore of arbuscular mycorrhizal fungi of the genus *Glomus*, Biometals 30 (2017) 559–564.
- [35] H. Etesami, B.R. Jeong, B.R. Glick, Contribution of arbuscular mycorrhizal fungi, phosphate-solubilizing bacteria, and silicon to P uptake by plant, Front. Plant Sci. 12 (2021) 1355.
- [36] R. Amiri, A. Nikbakht, N. Etemadi, M.R. Sabzalian, Nutritional status, essential oil changes and water-use efficiency of Rose geranium in response to arbuscular mycorrhizal fungi and water deficiency stress, Symbiosis 73 (2017) 15–25.
- [37] A.A. Meharg, J.W.G. Cairney, Ectomycorrhizas: extending the capabilities of rhizosphere remediation, Soil Biol. Biochem. 32 (2000) 1475–1484.
- [38] Y. Wang, M. Wang, Y. Li, A. Wu, J. Huang, Effects of arbuscular mycorrhizal fungi on growth and nitrogen uptake of *Chrysanthemum morifolium* under salt stress, PLoS One 13 (4) (2018) 1–14.
- [39] F. Martin, S. Duplessis, F. Ditengou, H. Lagrange, C. Voiblet, F. Lapeyrie, Developmental cross talking in the ectomychorrhizal symbiosis: signals and communication genes, New Phytol. 151 (2001) 145–154.
- [40] J. Lin, Y. Wang, S. Sun, C. Mu, X. Yan, Effects of arbuscular mycorrhizal fungi on the growth, photosynthesis and photosynthetic pigments of *Leymus Chinensis* seedlings under salt-alkali stress and nitrogen deposition, Sci. Total Environ. 576 (2017) 234–241.
- [41] A. Hashem, A.A. Alqarawi, R. Radhakrishnan, A.B.F. Al-Arjani, H.A. Aldehaish, D. Egamberdieva, E.F. Abd Allah, Arbuscular mycorrhizal fungi regulate the oxidative system, hormones and ionic equilibrium to trigger salt stress tolerance in Cucumis sativus L, Saudi J. Biol. Sci. 25 (2018) 1102–1114.
- [42] Q.S. Wu, Y.N. Zou, Arbuscular mycorrhizal fungi and tolerance of drought stress in plants, in: Q.S. Wu, eds.,

Arbuscular Mycorrhizas and Stress Tolerance of Plants, Springer, Singapore. 2017, pp. 25-41.

- [43] C. Kong, M. Camps-Arbestain, B. Clothier, P. Bishop, F.M. Vázquez, Use of either pumice or willow-based biochar amendments to decrease soil salinity under arid conditions, Environ. Technol. Innov. 24 (2021) 1–25.
- [44] S. Hussain, M. Shaukat, M. Ashraf, C. Zhu, Q. Jin, J. Zhang, Salinity stress in arid and semi-arid climates: effects and management in field crops, in: S. Hussain, eds., Climate Change and Agriculture, InTechOpen, London, UK. 2019, p. 198.
- [45] F. Zhan, B. Li, M. Jiang, X. Yue, Y. He, Y. Xia, Y. Wang, Arbuscular mycorrhizal fungi enhance antioxidant defense in the leaves and the retention of heavy metals in the roots of maize, Environ. Sci. Pollut. Res. 25 (2018) 24338–24347.
- [46] D. Pavithra, N. Yapa, Arbuscular mycorrhizal fungi inoculation enhances drought stress tolerance of plants, Groundw. Sustain. Dev. 7 (2018) 490–494.
- [47] N. Goicoechea, M.C. Antolín, Increased nutritional value in food crops, Microb. Biotechnol. 10 (2017) 1004–1007.
- [48] S. Ouledali, M. Ennajeh, A. Zrig, S. Gianinazzi, H. Khemira, Estimating the contribution of arbuscular mycorrhizal fungi to drought tolerance of potted olive trees (Olea europaea), Acta Physiol. Plant. 40 (2018) 1–13.
- [49] F. Zhang, J.D. He, Q.D. Ni, Q.S. Wu, Y.N. Zou, Enhancement of drought tolerance in trifoliate orange by mycorrhiza: changes in root sucrose and proline metabolisms, Not. Bot, Horti Agrobot 46 (2018) 270–276.
- [50] H. Lambers, F. Albornoz, L. Kotula, E. Laliberté, K. Ranathunge, F.P. Teste, G. Zemunik, How belowground interactions contribute to the coexistence of mycorrhizal and non-mycorrhizal species in severely phosphorusimpoverished hyperdiverse ecosystems, Plant Soil 424 (2018) 11–33.
- [51] S.G. Zardak, M.M. Dehnavi, A. Salehi, M. Gholamhoseini, Effects of using arbuscular mycorrhizal fungi to alleviate drought stress on the physiological traits and essential oil yield of fennel, Rhizosphere 6 (2018) 31–38.
- [52] D. Huang, M. Ma, Q. Wang, M. Zhang, G. Jing, C. Li, F. Ma, Arbuscular mycorrhizal fungi enhanced drought resistance in apple by regulating genes in the MAPK pathway, Plant Physiol. Biochem. 149 (2020) 245–255.
- [53] L. Tedersoo, M. Bahram, Mycorrhizal types differ in ecophysiology and alter plant nutrition and soil processes, Biol. Rev. Camb. Phil. Soc. 94 (2019) 1857–1880.
- [54] M. Semchenko, J.W. Leff, Y.M. Lozano, S. Saar, J. Davison, A. Wilkinson, B.G. Jackson, W.J. Pritchard, J.R. De Long, S. Oakley, K.E. Mason, N.J. Ostle, E.M. Baggs, D. Johnson, N. Fierer, R.D. Bardgett, Fungal diversity regulates plant-soil feedbacks in temperate grassland, Sci. Adv. 4 (11) (2018) 1–9.
- [55] M. Liang, D. Johnson, D. Burslem, S. Yu, M. Fang, J.D. Taylor, A.F.S. Taylor, T. Helgason, X. Liu, Soil fungal networks maintain local dominance of ectomycorrhizal trees, Nat. Commun. 11 (2020) 1–7.
- [56] S. van der Linde, L.M. Suz, C.D.L. Orme, F. Cox, H. Andreae, E. Asi, B. Atkinson, S. Benham, C. Carroll, N. Cools, B. De Vos, H.-P. Dietrich, J. Eichhorn, J. Gehrmann, T. Grebenc, H.S. Gweon, K. Hansen, F. Jacob, F. Kristöfel, P. Lech, M. Manninger, J. Martin, H. Meesenburg, P. Merilä, M. Nicolas, P. Pavlenda, P. Rautio, M. Schaub, H.-W. Schröck, W. Seidling, V. Šrámek, A. Thimonier, I.M. Thomsen, H. Titeux, E. Vanguelova, A. Verstraeten, L. Vesterdal, P. Waldner, S. Wijk, Y. Zhang, D. Žlindra, M.I. Bidartondo, Environment and host as large-scale controls of ectomycorrhizal fungi, Nature 558 (2018) 243–248.
- [57] A.A. Arpanahi, M. Feizian, G. Mehdipourian, D.N. Khojasteh, Arbuscular mycorrhizal fungi inoculation improve essential oil and physiological parameters and nutritional values of *Thymus daenensis* Celak and *Thymus vulgaris* L. under normal and drought stress conditions, Eur. J. Soil Biol. 100 (2020) 1–18.

- [58] M. Khalloufi, C. Martínez-Andújar, M. Lachaâl, N. Karray-Bouraoui, F. Pérez-Alfocea, A. Albacete, The interaction between foliar GA3 application and arbuscular mycorrhizal fungi inoculation improves growth in salinized tomato (*Solanum Lycopersicum* L.) plants by modifying the hormonal balance, J. Plant Physiol. 214 (2017) 134–144.
- [59] C. Averill, J.M. Bhatnagar, M.C. Dietze, W.D. Pearse, S.N. Kivlin, Global imprint of mycorrhizal fungi on wholeplant nutrient economics, Proc. Natl. Acad. Sci. USA 116 (2019) 23163–23168.
- [60] F. Zhang, M. Liu, Y. Li, Y. Che, Y. Xiao, Effects of arbuscular mycorrhizal fungi, biochar and cadmium on the yield and element uptake of Medicago sativa, Sci. Total Environ. 655 (2019) 1150–1158.
- [61] X.W. Chen, L. Wu, N. Luo, C.H. Mo, M.H. Wong, H. Li, Arbuscular mycorrhizal fungi and the associated bacterial community influence the uptake of cadmium in rice, Geoderma 337 (2019) 749–757.
- [62] A.H. Baghaie, F. Aghili, R. Jafarinia, Soil-indigenous arbuscular mycorrhizal fungi and zeolite addition to soil synergistically increase grain yield and reduce cadmium uptake of bread wheat (through improved nitrogen and phosphorus nutrition and immobilization of Cd in roots), Environ. Sci. Pollut. Res. 26 (2019) 30794–30807.
- [63] R.E. Abdelhameed, R.A. Metwally, Alleviation of cadmium stress by arbuscular mycorrhizal symbiosis, Int. J. Phytoremediation 21 (2019) 663–671.
- [64] M.G. Midgley, E. Brzostek, R.P. Phillips, Decay rates of leaf litters from arbuscular mycorrhizal trees are more sensitive to soil effects than litters from ectomycorrhizal trees, J. Ecol. 103 (2015) 1454–1463.
- [65] J.T. Wu, L. Wang, L. Zhao, X.C. Huang, F. Ma, Arbuscular mycorrhizal fungi affect growth and photosynthesis of *Phragmites australis* (Cav.) Trin Ex. Steudel under copper stress, Plant Biol. 22 (2020) 62–69.
- [66] Y. Ma, M. Rajkumar, R.S. Oliveira, C. Zhang, H. Freitas, Potential of plant beneficial bacteria and arbuscular mycorrhizal fungi in phytoremediation of metal-contaminated saline soils, J. Hazard Mater. 379 (2019) 1–15.
- [67] F. Lutzoni, M.D. Nowak, M.E. Alfaro, V. Reeb, J. Miadlikowska, M. Krug, A.E. Arnold, L.A. Lewis, D.L. Swofford, D. Hibbett, K. Hilu, T.Y. James, D. Quandt, S. Magallon, Contemporaneous radiations of fungi and plants linked to symbiosis, Nat. Commun. 9 (2018) 1–11.
- [68] M. Debeljak, J.T. van Elteren, A. Spruk, A. Izmer, F. Vanhaecke, K. Vogel-Mikuš, The role of arbuscular mycorrhiza in mercury and mineral nutrient uptake in maize, Chemosphere 212 (2018) 1076–1084.
- [69] F.M. Martin, S. Uroz, D.G. Barker, Ancestral alliances: plant mutualistic symbioses with fungi and bacteria, Science 356 (2017) 1–11.
- [70] N. Garg, S. Singh, Arbuscular mycorrhiza Rhizophagus irregularis and silicon modulate growth, proline biosynthesis and yield in *Cajanus cajan* L. Millsp. (Pigeonpea) genotypes under cadmium and zinc stress, J. Plant Growth Regul. 37 (2018) 46–63.
- [71] B. Pasbani, A. Salimi, N. Aliasgharzad, R. Hajiboland, Colonization with arbuscular mycorrhizal fungi mitigates cold stress through the improvement of antioxidant defense and accumulation of protecting molecules in eggplants, Sci. Hortic. 272 (2020) 1–22.
- [72] M. Atieno, L. Herrmann, H.T. Nguyen, H.T. Phan, N.K. Nguyen, P. Srean, M.M. Than, R. Zhiyong, P. Tittabutr, A. Shutsrirung, Assessment of biofertilizer use for sustainable agriculture in the Great Mekong Region, J. Environ. Manag. 275 (2020) 1–20.
- [73] J. Ma, W. Wang, J. Yang, S. Qin, Y. Yang, C. Sun, G. Pei, M. Zeeshan, H. Liao, L. Liu, Mycorrhizal symbiosis promotes the nutrient content accumulation and affects the root exudates in maize, BMC Plant Biol. 22 (2022) 1–13.
- [74] M. Kisiriko, M. Anastasiadi, L. Terry, A. Yasri, M.H. Beale, J.L. Ward, Phenolics from medicinal and aromatic plants:

characterization and potential as biostimulants and bioprotectants, Molecules 26 (2021) 1–37.

- [75] S. Basiru, H.P. Mwanza, M. Hijri, Analysis of arbuscular mycorrhizal fungal inoculant benchmarks, Microorganisms 9 (2021) 1–18.
- [76] N. Begum, C. Qin, M.A. Ahanger, S. Raza, M.I. Khan, M. Ashraf, N. Ahmed, L. Zhang, Role of arbuscular mycorrhizal fungi in plant growth regulation: implications in abiotic stress tolerance, Front. Plant Sci. 10 (2019) 1–15.
- [77] M. Amani Machiani, A. Javanmard, M.R. Morshedloo, M. Janmohammadi, F. Maggi, *Funneliformis mosseae* application improves the oil quantity and quality and ecophysiological characteristics of soybean (*Glycine max* L.) under water stress conditions, J. Soil Sci. Plant Nutr. 21 (2021) 3076–3090.
- [78] A. Genre, L. Lanfranco, S. Perotto, P. Bonfante, Unique and common traits in mycorrhizal symbioses, Nat. Rev. Microbiol. 18 (2020) 649-660.
- [79] M. Amani Machiani, A. Javanmard, M.R. Morshedloo, A. Aghaee, F. Maggi, Funneliformis mosseae inoculation under water deficit stress improves the yield and phytochemical characteristics of thyme in intercropping with soybean, Sci. Rep. 11 (2021) 1–13.
- [80] B. Mosse, Plant growth responses to vesicular-arbuscular mycorrhiza, New Phytol. 72 (1973) 127–136.
- [81] A.F. Fall, G. Nakabonge, J. Ssekandi, H. Founoune-Mboup, S.O. Apori, A. Ndiaye, A. Badji, K. Ngom, Roles of arbuscular mycorrhizal fungi on soil fertility: contribution in the improvement of physical, chemical, and biological properties of the soil, Front. Fungal Biol. 3 (2022) 1–11.
- [82] N. Golubkina, L. Krivenkov, A. Sekara, V. Vasileva, A. Tallarita, Prospects of arbuscular mycorrhizal fungi utilization in production of Allium plants, Plants 9 (2020) 1–16.
- [83] A. Ostadi, A. Javanmard, M. Amani Machiani, A. Sadeghpour, F. Maggi, M. Nouraein, M.R. Morshedloo, C. Hano, J.M. Lorenzo, Co-application of TiO2 nanoparticles and arbuscular mycorrhizal fungi improves essential oil quantity and quality of sage (*Salvia officinalis* L.) in drought stress conditions, Plants 11 (2022) 1–22.
- [84] P. Rozpądek, M. Rąpała-Kozik, K. Wężowicz, A. Grandin, S. Karlsson, Arbuscular mycorrhiza improves yield and nutritional properties of onion (*Allium cepa*), Plant Physiol. Biochem. 107 (2016) 264–272.
- [85] M.J. Harrison, Signaling in the arbuscular mycorrhizal symbiosis, Annu. Rev. Microbiol. 59 (2005) 19–42.
- [86] J. Li, B. Meng, H. Chai, X. Yang, W. Song, S. Li, A. Lu, T. Zhang, W. Sun, Arbuscular mycorrhizal fungi alleviate drought stress in C3 (*Leymus chinensis*) and C4 (*Hemarthria altissima*) grasses via altering antioxidant enzyme activities and photosynthesis, Front. Plant Sci. 10 (2019) 1–12.
- [87] M. Parniske, Molecular genetics of the arbuscular mychorrhiza symbiosis, Curr. Opin. Plant Biol. 7 (2004) 414–421.
- [88] D. Tagu, F. Lapeyrie, F. Martin, The ectomycorrhizal symbiosis: genetics and development, Plant Soil 244 (2002) 97–105.
- [89] S. Kaur, V. Suseela, Unraveling arbuscular mycorrhizainduced changes in plant primary and secondary metabolome, Metabolites 10 (2020) 1–30.
- [90] M.L. Goddard, L. Belval, I.R. Martin, L. Roth, H. Laloue, L. Deglène-Benbrahim, L. Valat, C. Bertsch, J. Chong, Arbuscular mycorrhizal symbiosis triggers major changes in primary metabolism together with modification of defense responses and signaling in both roots and leaves of *Vitis vinifera*, Front. Plant Sci. 12 (2021) 1–30.
- [91] F. Eulenstein, M. Tauschke, A. Behrendt, J. Monk, U. Schindler, M.A. Lana, S. Monk, The application of mycorrhizal fungi and organic fertilisers in horticultural potting soils to improve water use efficiency of crops, Horticulturae 3 (2017) 1–8.
- [92] S. Rashidi, A.R. Yousefi, M. Pouryousef, N. Goicoechea, Effect of arbuscular mycorrhizal fungi on the accumulation of secondary metabolites in roots and reproductive organs

of Solanum nigrum, Digitaria sanguinalis and Ipomoea purpurea, Chem. Biol. Technol. Agric. 9 (2022) 23-34.

- [93] A.B.F. Al-Arjani, A. Hashem, E.F. Abd\_Allah, Arbuscular mycorrhizal fungi modulates dynamics tolerance expression to mitigate drought stress in *Ephedra foliata* boiss, Saudi J. Biol. Sci. 27 (2020) 380–394.
- [94] E. Gamalero, G. Berta, N. Massa, B.R. Glick, G. Lingua, Synergistic interactions between the ACC deaminase-producing bacterium Pseudomonas putida UW4 and the AM fungus Gigaspora rosea positively affect cucumber plant growth, FEMS Microbiol. Ecol. 64 (2008) 459–467.
- [95] N. Moradtalab, R. Hajiboland, N. Aliasgharzad, T.E. Hartmann, G. Neumann, Silicon and the association with an ArbuscularMycorrhizal fungus (*rhizophagus clarus*) mitigate the adverse effects of drought stress on strawberry, Agronomy 9 (2019) 1–20.
- [96] J. Liu, X. Liu, Q. Zhang, S. Li, Y. Sun, W. Lu, C. Ma, Response of alfalfa growth to arbuscular mycorrhizal fungi and phosphate-solubilizing bacteria under different phosphorus application levels, Amb. Express 10 (2020) 200–213.
- [97] S. Gupta, S.D. Thokchom, R. Kapoor, Arbuscular mycorrhiza improves photosynthesis and restores alteration in sugar metabolism in *Triticum aestivum* L. Grown in arsenic contaminated soil, Front. Plant Sci. 12 (2021) 1–16.
- [98] R.A. Metwally, S.A. Soliman, A.A.H. Abdel Latef, R.E. Abdelhameed, The individual and interactive role of arbuscular mycorrhizal fungi and *Trichoderma viride* on growth, protein content, amino acids fractionation, and phosphatases enzyme activities of onion plants amended with fish waste, Ecotoxicol. Environ. Saf. 214 (2021) 1–13.
- [99] O.Y. Shtark, R.K. Puzanskiy, G.S. Avdeeva, V.V. Yemelyanov, M.S. Kliukova, A.L. Shavarda, A.A. Kirpichnikova, A.M. Afonin, I.A. Tikhonovich, V.A. Zhukov, Metabolic alterations in pea leaves and roots during arbuscular mycorrhiza development, PeerJ Comput. Sci. 7 (2020) 1–33.
- [100] A. Berruti, E. Lumini, R. balestrini, V. Bianciotto, Wild Camellia japonica specimens in the Shimane prefecture (Japan) host previously undescribed AMF diversity, Appl. Soil Ecol. 115 (2017) 10–18.
- [101] A. Andrino, G. Guggenberger, S. Kernchen, R. Mikutta, L. Sauheitl, J. Boy, Production of organic acids by arbuscular mycorrhizal fungi and their contribution in the mobilization of phosphorus bound to iron oxides, Front. Plant Sci. 12 (2021) 1–13.
- [102] E. Rezaei-Chiyaneh, M.L. Battaglia, A. Sadeghpour, F. Shokrani, A.D.M. Nasab, M.A. Raza, M. von Cossel, Optimizing intercropping systems of black cumin (*Nigella sativa* L.) and fenugreek (*Trigonella foenum-graecum* L.) through inoculation with bacteria and mycorrhizal fungi, Adv. Sustain. Syst. 5 (9) (2021) 1–14.
- [103] J.H. Graham, Assessing cost of arbuscular mycorrhizal symbiosis in agroecosystems, in: G.K. Podila, D.D. Douds Jr., eds., Current Advances in Mycorrhizal Research, APS Press, St. Paul, NM. 2000, pp. 127–140.
- [104] R.T. Koide, M.D. Goff, I.A. Dickie, Component growth efficiencies of mycorrhizal and nonmycorrhizal plants, New Phytol. 148 (2000) 163–168.
- [105] A.K. Srivastava, S. Singh, R.A. Marathe, Organic citrus, soil fertility and plant nutrition, J. Sustain. Agric. 19 (2000) 5–29.
- [106] A. Mohammad, B. Mitra, A.G. Khan, Effects of sheared-root inoculurn of *Glomus intraradices* on wheat grown at different phosphorus levels in the field, Agric. Ecosyst. Environ. 103 (2004) 245–249.
- [107] P. Gosling, A. Hodge, G. Goodlass, G.D. Bending, Arbuscular mycorrhizal fungi and organic farming, Agric. Ecosyst. Environ. 113 (2006) 17–35.
- [108] V.A. Borowicz, Do arbuscular mycorrhizal fungi alter plant-pathogen relations? Ecology 82 (2001) 3057-3068.
  [109] N. Karagiannidis, F. Bletsos, N. Stavropoulos, Effect of
- [109] N. Karagiannidis, F. Bletsos, N. Stavropoulos, Effect of Verticillium wilt (Verticillium dahliae Kleb.) and mycorrhiza (Glomus mosseae) on root colonization, growth and nutrient

uptake in tomato and eggplant seedlings, Sci. Hortic. 94 (2002) 145-156.

- [110] J.M. Whipps, Prospects and limitations for mycorrhizas in biocontrol of root pathogens, Can. J. Bot. 82 (2004) 1198–1227.
- [111] C. Azcon-Aguilar, J.M. Barea, Arbuscular mycorrhizas and biocontrol of soil-borne plant pathogens, an overview of the biological mechanisms involved, Mycorrhiza 6 (1996) 457–464.
- [112] R.M. Auge, Arbuscular mycorrhizae and soil/plant water relations, Can. J. Soil Sci. 84 (2004) 373–381.
- [113] G.N. Al-Karaki, R. Hammad, M. Rusan, Response of two tomato cultivars differing in salt tolerance to inoculation

with mycorrhizal fungi under salt stress, Mycorrhiza 11 (2001) 43-47.

- [114] G. Feng, F.S. Zhang, X.L. Li, C.Y. Tian, C. Tang, Z. Rengel, Improved tolerance of maize plants to salt stress by arbuscular mycorrhiza is related to higher accumulation of soluble sugars in roots, Mycorrhiza 12 (2002) 185–190.
- [115] I. Jakobsen, L. Rosendahl, Carbon flow into soil and external hyphae from roots of mycorrhizal cucumber plants, New Phytol. 115 (1990) 77–83.
- [116] M.C. Rillig, C.B. Field, M.F. Allen, Soil biota responses to longterm atmospheric CO<sub>2</sub> enrichment in two California annual grasslands, Oecologia 119 (1999) 572–577.