

Karbala International Journal of Modern Science

Volume 9 | Issue 3

Article 12

Mutation-Induced Changes in the Stability, B-Cell Epitope, and Antigenicity of the Sars-Cov-2 Variant Spike Protein: A Comparative Computational Stud

Nira Meirita Wijayanti Biology Department, Faculty of mathematics and Natural Sciences, Brawijaya University, Malang, Indonesia

Muhammad Hermawan Widyananda Biology Department, Faculty of mathematics and Natural Sciences, Brawijaya University, Malang, Indonesia

Lailil Muflikhah Department of Informatics Engineering, Faculty of Computer Science, Brawijaya University, Malang, Indonesia

Nashi Widodo Biology Department, Faculty of mathematics and Natural Sciences, Brawijaya University, Malang, Indonesia, widodo@ub.ac.id

Follow this and additional works at: https://kijoms.uokerbala.edu.ig/home

Part of the Bioinformatics Commons, Biomedical Informatics Commons, Other Immunology and Infectious Disease Commons, Virology Commons, and the Virus Diseases Commons

Recommended Citation

Wijayanti, Nira Meirita; Widyananda, Muhammad Hermawan; Muflikhah, Lailil; and Widodo, Nashi (2023) "Mutation-Induced Changes in the Stability, B-Cell Epitope, and Antigenicity of the Sars-Cov-2 Variant Spike Protein: A Comparative Computational Stud," *Karbala International Journal of Modern Science*: Vol. 9 : Iss. 3 , Article 12. Available at: https://doi.org/10.33640/2405-609X.3311

This Research Paper is brought to you for free and open access by Karbala International Journal of Modern Science. It has been accepted for inclusion in Karbala International Journal of Modern Science by an authorized editor of Karbala International Journal of Modern Science. For more information, please contact abdulateef1962@gmail.com.



Mutation-Induced Changes in the Stability, B-Cell Epitope, and Antigenicity of the Sars-Cov-2 Variant Spike Protein: A Comparative Computational Stud

Abstract

The spike (S) protein is a major antigenicity site that targets neutralizing antibodies and drugs. The growing number of S protein mutations has become a severe problem for developing effective vaccines. Here, we investigated four severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) variants that were the most infectious and widespread during the COVID-19 pandemic to determine the trends and patterns of mutation-induced changes in the stability, B-cell epitope, and antigenicity of the SARS-CoV-2 S protein. The data showed that the Beta and Gamma variants had three mutations on the receptor-binding domain (RBD), which is the specific site on the S protein for angiotensin-converting enzyme 2 (hACE2) binding. The Delta variant had only two mutations, whereas the Omicron variant had 15 mutations on the RBD. The results showed that the stability of the S protein varied and depended on the mutation type and that Gamma and Omicron are the most stable of the four variants analyzed. The S protein-hACE2 complexes of the Beta and Gamma variants were relatively stable after 20 ns of simulation compared with those of the Delta and Omicron variants. We predicted that the B-cell epitopes of the mutant S protein would be different from those of the wildtype. Moreover, the antigenicity of Omicron changed drastically compared with that of the other variants. Bioinformatics analysis and a molecular dynamic simulation revealed that the mutations affected the stability of the S protein. A large number of mutations do not always stabilize the S protein. Mutations in Omicron significantly altered the B-cell epitope and antigenicity, which decreased vaccine effectiveness. These findings provide insights into SARS-CoV-2 evolution for vaccine development.

Keywords

Antigenicity; B-cell epitope; Mutation; SARS-CoV-2; S Protein

Creative Commons License



This work is licensed under a Creative Commons Attribution-Noncommercial-No Derivative Works 4.0 License.

Mutation-induced Changes in the Stability, B-cell Epitope, and Antigenicity of the Sars-Cov-2 Variant Spike Protein: A Comparative Computational Study

Nira M. Wijayanti^a, Muhammad H. Widyananda^a, Lailil Muflikhah^b, Nashi Widodo^{a,*}

^a Biology Department, Faculty of Mathematics and Natural Sciences, Brawijaya University, Malang, Indonesia

^b Department of Informatics Engineering, Faculty of Computer Science, Brawijaya University, Malang, Indonesia

Abstract

The spike (S) protein is a major antigenicity site that targets neutralizing antibodies and drugs. The growing number of S protein mutations has become a severe problem for developing effective vaccines. Here, we investigated four severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) variants that were the most infectious and widespread during the COVID-19 pandemic to determine the trends and patterns of mutation-induced changes in the stability, B-cell epitope, and antigenicity of the SARS-CoV-2 S protein. The data showed that the Beta and Gamma variants had three mutations on the receptor-binding domain (RBD), which is the specific site on the S protein for angiotensin-converting enzyme 2 (hACE2) binding. The Delta variant had only two mutations, whereas the Omicron variant had 15 mutations on the RBD. The results showed that the stability of the S protein varied and depended on the mutation type and that Gamma and Omicron are the most stable of the four variants analyzed. The S protein-hACE2 complexes of the Beta and Gamma variants were relatively stable after 20 ns of simulation compared with those of the Delta and Omicron variants. We predicted that the B-cell epitopes of the mutant S protein would be different from those of the wildtype. Moreover, the antigenicity of Omicron changed drastically compared with that of the other variants. Bioinformatics analysis and a molecular dynamic simulation revealed that the mutations affected the stability of the S protein. A large number of mutations do not always stabilize the S protein. Mutations in Omicron significantly altered the B-cell epitope and antigenicity, which decreased vaccine effectiveness. These findings provide insights into SARS-CoV-2 evolution for vaccine development.

Keywords: Antigenicity, B-cell epitope, Mutation, SARS-CoV-2, S protein

1. Introduction

S evere acute respiratory syndrome coronavirus 2 (SARS-CoV-2), the causative agent of the coronavirus disease (COVID-19), is a highly mutational RNA virus, as it lacks a proofreading mechanism. This virus continuously undergoes deletion, insertion, and genetic recombination that change critical amino acids in its genome. These amino acid changes benefit adaptation and evolution of the coronavirus. This continual series of mutations has led to a vast diversity of variants of the virus from

the original Wuhan sequence [1-3]. In February 2022, several variants of concern, including Gamma (P.1), Beta (B.1.351), Omicron (B.1.529), and Delta (B.1.617), were identified. The continuously mutating SARS-CoV-2 spike (S) protein is a severe problem for developing effective vaccines against COVID-19.

The S protein is a part of the structure that plays a unique and crucial function during the early stage of infection to angiotensin-converting enzyme 2 (hACE2), the target-cell host receptor. As a target of antibodies and drugs, the S protein is a major

Received 13 November 2022; revised 3 July 2023; accepted 7 July 2023. Available online 12 August 2023

* Corresponding author. E-mail address: widodo@ub.ac.id (N. Widodo). antigenicity site. Mutations in the S protein have become the primary concern of many studies because the S protein is an important target for neutralizing antibodies [4,5]. Each variant demonstrates a different mutation on its S protein. Mutations affect the stability and binding expression of S-hACE2; however, no study has compared the mutational effect of the SARS-CoV-2 S protein variants of concern. Therefore, in this study, we investigated four SARS-CoV-2 variants that were the most infectious and widespread during the pandemic (Beta, Delta, Gamma, and Omicron) to detect trends and patterns of how mutations in the S protein affect interactions with ACE2 due to structural and functional changes in the S protein. We also compared the effect of mutations of the B-cell epitopes and antigenicity of the four SARS-CoV-2 variants.

In this study, we investigated the mutational effect, epitope, and antigenicity of the four abovementioned SARS-CoV-2 variants using bioinformatics analysis and molecular dynamic (MD) simulation and found that the stability of the S protein was affected by the mutations. The results of the MD simulations showed that Beta and Gamma were relatively more stable than Delta and Omicron, revealing that an increase in the number of mutations does not always correlate with stabilization of the S protein. Accumulated mutations on Omicron significantly changed its B-cell epitope and antigenicity, thereby decreasing the effectiveness of vaccines. These findings provide a more comprehensive understanding of SARS-CoV-2 adaptation and evolution.

2. Materials and methods

2.1. Data mining

The S-hACE2 crystalline structure was obtained from the Protein Data Bank (http://www.pdb.org). The

entire genome of the SARS-CoV-2 spike glycoprotein structure was constructed using the Swiss Model Homolog on the ExPASy web server (swissmodel.expasy. org). Sequences were retrieved from all selected variants of the protein using Biovia Discovery Studio.

2.2. Mutation collection

We introduced a mutation to the whole genome monomer S protein and the S protein complex, containing hACE2, by substituting amino acids based on the mutation list retrieved from the literature [10–12] to obtain the desired SARS-CoV-2 variants (Beta, Gamma, Delta, and Omicron) using the FOLDX5 plugin on YASARA (Table 1) (viralzone.expasy.org). Due to application limitations, the mutations introduced were limited to amino acid substitutions (i.e., no deletions or insertions). Amino acid substitutions were introduced into the S protein monomer throughout the genome, not just on the receptor-binding domain (RBD). The mutations were only introduced on the RBD of the S protein—hACE2 complex.

2.3. Protein stability calculation

The stability of the SARS-CoV-2 S protein and the S–hACE2 protein complex was predicted by the folding free energy change ($\Delta\Delta G$) between the wild-type (WT) and the mutant S protein structure. FoldX5 is a popular tool to assess the effect of a mutation on the folding and dynamics of nucleic acids and proteins [6].

The folding free energy change was calculated as follows [7]:

$\Delta\Delta G$ (stability) = ΔG (folding)mutant - ΔG (folding)WT

A negative G value indicates that the mutation stabilizes the protein, whereas a positive G value indicates that the mutation destabilizes the protein.

Table 1. Mutation sites in the RBD S protein.

Origin	First Reported	Lineage	Mutation on S Protein	Ref
Beta	September 2020	B.1.351	K417N; E484K; N501Y; D614G; A701V	[10]
(South Africa)				
Gamma	January 2021	P.1	L18F; T20N; P26S; D138Y; R190S; K417T;	[10]
(Japan & Brazil)			E484K; N501Y; D614G; H655Y; T1027I	
Delta	October 2021	B.1.617	T19R, G142D, Δ156–157, R158G, Δ213–214,	[11]
(India)			L452R, T478K, D614G, P681R, D950N	
Omicron	November 2021	B.1.529	A67V; Δ69-70del; T95I; G142D; Δ143del,	[12]
(South Africa)			G143D, Δ144-145del, Δ211del; L212I; ins214EPE;	
			G339D; S371L; S373P; S375F: K417N; N440K; G446S;	
			S477N; T478K; E484A; Q493R; G496S; Q498R; N501Y;	
			Y505H; T547K; D614G; H655Y; N679K; P681H; N764K;	
			D796Y; N856K; Q954H; N969K; L981F	
			D7901, N030K, Q93411, N909K, L9011	

2.4. Molecular dynamics simulation

The effect of each mutation on the S protein was assessed, and the structural stability was predicted using MD. The MD analysis was based on the analysis of the physical movement of atoms and molecules during a simulation. The MD analysis was performed using YASARA with 20 ns of simulation time. The MD simulations were conducted on parameters that were adapted to cellular conditions. The system was neutralized with 0.9% NaCl and 1 atm of pressure while maintaining a pH of 7.4. The temperature was set to 298 K. The simulation was conducted with the md_runfast program in the autosave setting every 25 ps for up to 800 simulations.

2.5. Predicting the B-cell epitopes and antigenicity

The B-cell epitope was predicted using BepiPred-1.0 Linear Epitope Prediction at the Iedb.org website. BepiPred is a machine learning-based method that predicts epitopes using the propensity scale and hidden Markov models [6,8]. The WT and SARS-CoV-2 mutant S protein antigenicity was predicted at the iedb.org website using the Kolaskar and Tongaonkar antigenicity scales [9].

3. Results and discussion

The SARS-CoV-2 variants exhibited different mutations on the S protein. Among the four variants analyzed, the Beta variant had the fewest mutations, with three mutations on the RBD and three mutations on the S sequence. The Gamma variant had 3 mutations on the RBD and 11 on the S sequence. The Delta variant had two mutations on the RBD and seven on the S sequence. The Omicron variant had 15 mutations on the RBD and 30 on the S protein (Table 1).

3.1. Mutations that altered the S protein structure and increased the protein stability

The S protein is important during the initial stages of virus infection. The S protein binds with hACE2 to enter the cell. Due to a strong correlation between structure and function, the structural stability of all mutant S proteins was analyzed to understand the effect of the mutation on protein stability. The S protein was stabilized after one mutation (Table 2), and the mutants had different degrees of stability depending on the mutation. Gamma and Omicron were the most stable variants.

The root-mean-square deviation (RMSD) backbone of the four SARS-CoV-2 variants showed the effect of mutation on the structural stability of the

able 2. Ar	чіпо ас.	id sequ	ence ch	anges ii	n the w	ildtype	and SA	RS-Col	7-2 mut	ant.														
/ariant	Posit	ion of	Aminc	Acid																		ΔG	ΔG	ΔΔG
	18	19	20	26	67	95	138	142	158	190	212	339	371	373	375	417	440	14 6 4	52 4	. 42	478	wildtype	mutant	
Vildtype	Г	Г	н	Р	Α	Т	D	U	R	R	L	ß	s	s	s	K	Z	U U	. 1		г			
seta	L	Г	Г	Ь	A	Г	D	U	R	R	L	U	s	s	s	z	z	ר ט	. 1		н	750.06	749.52	-0.54
Jamma	H	Г	Z	s	A	Г	Y	IJ	R	s	L	ť	s	s	s	Т	z	ו ט	. 1		н	736.73	731.30	-5.43
Delta	Γ	Ч	Г	Ъ	A	Г	D	U	IJ	R	L	ť	s	s	s	Х	z	۳ ت	~		н	751.34	750.84	-0.50
Dmicron	L	Г	Г	Ь	>	I	D	D	R	R	I	D	L	Ь	ц	z	×	S	-	7	×	707.44	704.21	-3.23
	484	493	496	498	501	505	547	614	655	679	681	701	764	796	856	950	954	5 696	81 1	027				
Vildtype	щ	0	U	0	z	٢	Г	D	Η	Z	Р	A	z	D	z	D	0	I Z	. 1	r .				
eta	Ч	0	U	0	۲	۲	Т	U	Η	z	Р	>	z	D	z	D	0	ı z	. 1	r .		750.06	749.52	-0.54
Jamma	Ч	0	U	0	¥	¥	Т	U	٢	z	Ъ	A	z	D	z	D	0	z	н . 1			736.73	731.30	-5.43
Delta	Щ	0	U	0	z	Y	Г	IJ	Η	z	R	A	z	D	z	z	0	z		F .		751.34	750.84	-0.50
Omicron	A	R	s	R	Y	Η	Х	ט	Y	ч	Η	A	¥	Y	X	D	H	X	[T.	r.,		707.44	704.21	-3.23

S protein. The RMSD analysis indicated that the WT and mutants had high RMSD values after 20 ns of simulation time, which may have been due to an unstable S protein monomer. To be stable, the S protein must be in the trimer state. The stability of the variants was compared with that of the WT for the structural stability analysis (Fig. 1).

The fluctuating trends were not different among the WT, Gamma, and Delta variants. Omicron had a lower RMSD value despite its instability. The RMSD analysis of Beta indicated relatively stable fluctuations, starting at 13 ns of simulation. Beta was more stable than the other variants.

The root-mean-square fluctuation (RMSF) depicts the flexibility of the amino acid residue under study over the simulation time [13]. The RMSF analysis showed that the WT and mutants had similar fluctuating trends in the protein residues. The protein fluctuated the least during the start of the simulation, increased toward the middle, and then decreased at the end. In contrast, Gamma displayed the greatest fluctuations in the region between Phe392 and Thr581, the region between Phe833 and Lys854, and the site between Leu959 and Leu1001 compared with the other strains. Beta displayed the least fluctuations compared with the other strains in the same region. This RMSF result supports the RMSD data, showing that the amino acids in a protein structurally contribute most to molecular motion.

Fig. 2 depicts the mutations in the S1-RBD of the SARS-CoV-2 variants. Beta and Gamma had similar mutations in the S1-RBD. The amino acids on the RBD of Beta and Gamma changed to E484K, N501Y, and K417N. The Delta variant only carried the L452R and T478K mutations. Omicron has recently evolved into many subvariants and has gained 15 mutations in its RBD.

The RMSD backbone of the S protein—hACE2 WT complex and mutant SARS-CoV-2 was also analyzed. The RMSD analysis revealed that Beta and Gamma had 2 Å values and relatively stable fluctuations after 20 ns of simulation, indicating that the S proteins of these variants were stable after binding to hACE2. Delta and Omicron had values exceeding 3 Å and were relatively unstable after 3 and 8 ns of simulation, indicating that the mutations disrupted the binding interactions between the S protein and hACE2.

3.2. Omicron antigenicity was altered drastically compared with the other variants

The B-cell epitope refers to the part of the antigen that binds to the antibody. The B-cell epitope is recognized by B-cell receptors (BCRs) and induces an antibody response [6]. Because the S protein



Fig. 1. Analysis of trajectory fluctuations in the molecular dynamics simulations of the SARS-CoV-2 wildtype and mutant S proteins. (A) Root-meansquare-deviation. (B) Root-mean-square fluctuation.





Fig. 2. Mutation mapping of the S-hACE2-SARS-CoV-2 variant complex. (A) Beta-Gamma. (B) Delta. (C) Omicron.

epitopes are important for triggering an immune response, presenting the S protein might be a rationale for several essential vaccines. This is a critical issue concerning the long-term effectiveness of newly developed vaccines. We predicted and compared the B-cell epitopes of the WT and mutant S proteins using a well-established predictive web server tool. The S protein mutants had different epitope variations, which resulted in differences in the efficacy of the antibodies and vaccines (Figs. 2A and 4A). The B-cell epitope profiles of the Gamma, Delta, and Omicron variant mutants were greater than the threshold value (0.5), indicating that these variants are easier to recognize using the BCR of infected cells compared with the WT and Beta variants.

A

F484

Antigenicity describes the capacity of a virus to bind to antibodies. Variations in the antigenicity at these sites are driven by neutralizing antibodies. Changes in mutation-induced antigenicity play a role in the effectiveness of vaccines [13–15]. We compared the antigenicity fluctuations of the Beta, Gamma, Delta, and Omicron variants. Each variant differed in terms of antigenicity. Omicron antigenicity was severely altered compared with that of the other variants (threshold value: 1.05) (Figs. 2B and 4B).

Multiple mutations have repeatedly occurred on the SARS-CoV-2 S protein, resulting in the current SARS-CoV-2 variants. To understand the trend in the mutations and the evolution of the structure of the four variants, we compared their protein stability, B-cell epitope, and antigenicity. The first case of Beta was detected in September 2020 in South Africa. Beta was linked with an increased hospitalization rate, immune escape, and death. The Beta variant exhibited no increase in transmissibility [16]. Beta became the most prevalent variant in the second wave of the pandemic after the Alpha variant. Besides D614G, the global mutations in SARS-CoV-2 are known to benefit the virus with faster transmission rates, higher affinity, and higher antigenicity [17]. Beta also gained the E484K and N501Y mutations, which enhanced its binding affinity to hACE2 compared with the WT [18].

The Gamma variant has a lower hospitalization risk than the Beta variant, which is associated with increased transmissibility and immune response escape. The Gamma variant shares some mutations with Beta (N501Y, E484K, and D614G) and has also



Fig. 3. Backbone fluctuations of the wildtype and mutant S protein—hACE2 complexes. The RMSD value indicates the dynamic of backbone movement of the complex of S protein with hACE2.



Fig. 4. Linear B-cell epitopes and antigenicity of the SARS-CoV-2 variants. (A) Linear B-cell epitope profile. (B) Antigenicity profile.

gained additional mutations (L18F and K417T). These mutations benefit the virus with evasion of antibody-mediated immunity, reinfection, and increased transmission rates [19].

The Delta variant gained the E484Q and L452R mutations, which are associated with increased hACE2 binding affinity to the RBD. Delta also has a P681R mutation on its S protein sequence, which enhances its immune escape ability. Omicron is the most recent variant circulating in the population and has accumulated the most mutations in its sequence (Fig. 4). Omicron has 30 mutations in the S protein sequence. Omicron has been linked to significantly lower hospitalization and mortality rates than previous variants [20,21].

These results agree with a previous report that suggested that Omicron reduces the neutralization activity of antibodies [22], but its infection fatality rate is lower. Nevertheless, Omicron has a three-fold higher transmissibility rate due to immune escape than previous variants based on South African findings. The mutations appear to alter the dynamics of the protein-protein interactions in the S sequence, which helps stabilize the S protein [18,23–25]. A stable S protein binds more effectively to the ACE2 receptor. A mutation might change the stability of the S protein differently depending on which amino acid undergoes the mutation. Mutations, particularly in the RBD, have various effects on protein stability and ACE2 binding [26]. Different amino acid mutations exert different effects.

Some mutations are associated with higher viral antigenicity [27]. Changes in antigenicity result in the changes in antibody's ability to recognize antigens and reduce the effectiveness of the previous vaccine. This could be the rationale behind the booster vaccine being a prerequisite to protect the population and minimize the effect of the SARS-CoV-2 variants. A vaccine developed for a specific variant may be ineffective or less effective in protecting against future evolving variants. Omicron has been reported to be less protective against infection with two and three doses of vaccination than Delta [28,29]. This phenomenon is mainly caused by the mutations in the S protein, especially on the RBD; Omicron has 30 on its S protein, of which half reside on the RBD [29-31]. This mutation rate is twice that of Delta.

The RMSD results of the S monomer and S protein—hACE2 complex revealed that the Beta and Gamma variants were more likely to be stable than the Delta and Omicron variants, indicating that increasing the number of mutations does not always result in the stabilization of the S protein (Fig. 3). The RMSF results support this finding.

4. Conclusion

Mutations affect the stability of the S protein. However, increasing the number of mutations does not always result in a stable S protein. Moreover, the mutations we induced had different effects on the epitope and antigenicity of each variant. The growing number of mutations in Omicron has significantly changed its antigenicity compared with that of the original Wuhan sequence and other mutant variants, potentially reducing vaccine effectiveness. Our research provides a more comprehensive understanding of the effects of mutation and the evolution of SARS-CoV-2 for long-term vaccine development. Our study should be used as preliminary screening only, as it has computational limitations. Laboratory experimental research is required in future studies.

Conflicts of interest

None of the authors declares any potential conflicts of interest regarding this study.

Acknowledgments

The authors thank the Bioinformatics and Computational Biology Laboratory, Department of Biology, Brawijaya University, for the facilities provided. The authors also thank the Faculty of Mathematics and Natural Sciences, Brawijaya University for the research funding of Professor Grant's scheme.

References

- [1] S.S. Boon, C. Xia, M.H. Wang, K.L. Yip, H.Y. Luk, S. Li, R.W.Y. Ng, C.K.C. Lai, P.K.S. Chan, Z. Chen, Temporalgeographical dispersion of SARS-CoV-2 spike glycoprotein variant lineages and their functional prediction using *in silico* approach, mBio 12 (2021) 1–19, https://doi.org/10.1128/ mBio.02687-21.
- [2] R. Khandia, S. Singhal, T. Alqahtani, M.A. Kamal, N.A. El-Shall, F. Nainu, P.A. Desingu, K. Dhama, Emergence of SARS-CoV-2 Omicron (B.1.1.529) variant, salient features, high global health concerns, and strategies to counter it amid the ongoing COVID-19 pandemic, Environ. Res. 209 (2022) 112816, https://doi.org/10.1016/j.envres.2022.112816.
- [3] B.O. Villoutreix, V. Čalvez, A.-G. Marcelin, A.-M. Khatib, Silico investigation of the new UK (B.1.1.7) and South African (501Y.V2) SARS-CoV-2 variants with a focus at the ACE2-spike RBD interface, Int. J. Mol. Sci. 22 (2021) 1695, https:// doi.org/10.3390/ijms22041695.
- [4] Y.D. Devi, H.B. Goswami, S. Konwar, C. Doley, A. Dolley, A. Devi, C. Chongtham, D. Dowerah, V. Biswa, L. Jamir, A. Kumar, S.S. Satapathy, S.K. Ray, R.C. Deka, R. Doley, M. Mandal, S. Das, C.S. Singh, P.P. Borah, P. Nath, N.D. Namsa, Immunoinformatics Mapping of Potential Epitopes in SARS-CoV-2 Structural Proteins, vol. 16, 2021, pp. 1–45, https://doi.org/10.1371/journal.pone.0258645.

- [5] C.S. Lupala, Y. Ye, H. Chen, X.D. Su, H. Liu, Mutations on RBD of SARS-CoV-2 Omicron variant result in stronger binding to the human ACE2 receptor, Biochem. Biophys. Res. Commun. 590 (2022) 34–41, https://doi.org/10.1016/ J.BBRC.2021.12.079.
- [6] J.L. Sanchez-Trincado, M. Gomez-Perosanz, P.A. Reche, Review Article Fundamentals and Methods for T-And B-Cell Epitope Prediction, 2017, 2017, pp. 1–14, https://doi.org/ 10.1155/2017/2680160.
- [7] S. Teng, A. Sobitan, R. Rhoades, D. Liu, Q. Tang, Systemic effects of missense mutations on SARS-CoV-2 spike glycoprotein stability and receptor-binding affinity, Brief. Bioinform. 22 (2021) 1239–1253, https://doi.org/10.1093/bib/bbaa233.
- [8] J. Larsen, O. Lund, M. Nielsen, Improved method for predicting linear B-cell epitopes, Immunome Res 2 (2006) 2, https://doi.org/10.1186/1745-7580-2-2.
- [9] A.S. Kolaskar, C. Tongaonkar, Kolaskar A Semi-empirical Method for the Prediction of Antigenic Determinants on protein.Pdf, Elsevier Sci. vol. 276 (1990) 172–174, https:// doi.org/10.1016/0014-5793(90)80535-q.
- [10] S.E. Galloway, P. Paul, D.R. MacCannell, M.A. Johansson, J.T. Brooks, A. MacNeil, R.B. Slayton, S. Tong, B.J. Silk, G.L. Armstrong, M. Biggerstaff, V.G. Dugan, Emergence of SARS-CoV-2 B.1.1.7 lineage — United States, 29 december 2020–12 january 2021, MMWR Morb. Mortal. Wkly. Rep. 70 (2021) 95–99, https://doi.org/10.15585/mmwr.mm7003e2.
- [11] S. Kumar, T.S. Thambiraja, K. Karuppanan, G. Subramaniam, Omicron and Delta variant of SARS-CoV-2: a comparative computational study of spike protein, J. Med. Virol. 94 (2022) 1641–1649, https://doi.org/10.1002/jmv.27526.
- [12] A. Kumar, A. Asghar, H.N. Singh, M.A. Faiq, S. Kumar, R.K. Narayan, G. Kumar, P. Dwivedi, C. Sahni, R.K. Jha, M. Kulandhasamy, P. Prasoon, K. Sesham, K. Kant, S.N. Pandey, An in silico analysis of early SARS-CoV-2 variant B.1.1.529 (Omicron) genomic sequences and their epidemiological correlates, medRxiv 529 (2021) 1–21, https:// doi.org/10.1101/2021.12.18.21267908.
- [13] D. Dey, P. Paul, S. Azad, M. Mazid, A. Khan, M. Sharif, M. Rahman, Molecular optimisation, docking, and dynamic simulation profiling of selective aromatic phytochemical ligands in blocking the SARS-CoV-2 S protein attachment to ACE2 receptor: an in silico approach of targeted drug designing, J. Adv. Vet. Anim. Res. 8 (2021) 1, https://doi.org/ 10.5455/javar.2021.h481.
- [14] R. Servín-Blanco, R. Zamora-Alvarado, G. Gevorkian, K. Manoutcharian, Antigenic variability: obstacles on the road to vaccines against traditionally difficult targets, Hum. Vacc. Immunother. 12 (2016) 2640–2648, https://doi.org/ 10.1080/21645515.2016.1191718.
- [15] J. Wu, J. Nie, L. Zhang, H. Song, Y. An, Z. Liang, J. Yang, R. Ding, S. Liu, Q. Li, T. Li, Z. Cui, M. Zhang, P. He, Y. Wang, X. Qu, Z. Hu, Q. Wang, W. Huang, The antigenicity of SARS-CoV-2 Delta variants aggregated 10 high-frequency mutations in RBD has not changed sufficiently to replace the current vaccine strain, Signal Transduct. Target. There. 7 (2022) 18, https://doi.org/10.1038/s41392-022-00874-7.
- [16] D. Frampton, T. Rampling, A. Cross, H. Bailey, J. Heaney, M. Byott, R. Scott, R. Sconza, J. Price, M. Margaritis, M. Bergstrom, M.J. Spyer, P.B. Miralhes, P. Grant, S. Kirk, C. Valerio, Z. Mangera, T. Prabhahar, J. Moreno-Cuesta, N. Arulkumaran, M. Singer, G.Y. Shin, E. Sanchez, S.M. Paraskevopoulou, D. Pillay, R.A. McKendry, M. Mirfenderesky, C.F. Houlihan, E. Nastouli, Genomic characteristics and clinical effect of the emergent SARS-CoV-2 B.1.1.7 lineage in London, UK: a whole-genome sequencing and hospital-based cohort study, Lancet Infect Dis 21 (2021) 1246–1256, https://doi.org/10.1016/S1473-3099(21)00170-5.
- 1246–1256, https://doi.org/10.1016/S1473-3099(21)00170-5.
 [17] J.A. Plante, Y. Liu, J. Liu, H. Xia, B.A. Johnson, K.G. Lokugamage, X. Zhang, A.E. Muruato, J. Zou, C.R. Fontes-Garfias, D. Mirchandani, D. Scharton, J.P. Bilello, Z. Ku, Z. An, B. Kalveram, A.N. Freiberg, V.D. Menachery, X. Xie, K.S. Plante, S.C. Weaver, P.Y. Shi, Spike mutation

D614G alters SARS-CoV-2 fitness, Nature 592 (2021) 116-121, https://doi.org/10.1038/s41586-020-2895-3.

- [18] I. Čelik, A. Khan, F. Martha Dwivany, D.-Q. Wei, T. Ekawati Tallei, Computational prediction of the effect of mutations in the receptor-binding domain on the interaction between SARS-CoV-2 and human ACE2, Mol. Divers. 1 (2022) 3, https://doi.org/10.1007/s11030-022-10392-x.
- [19] R. Cantón, P.D.L. Ramos, A. García-Botella, A. García-Lledó, J. Gómez-Pavón, J.G. Del Castillo, T. Hernández-Sampelayo, M.C. Martín-Delgado, F.J.M. Sánchez, M. Martínez-Sellés, J.M.M. García, S.M. Guillén, F. Rodríguez-Artalejo, J. Ruiz-Galiana, E. Bouza, New variants of SARS-CoV-2, Rev. Esp. Quimioter. 34 (2021) 419–428, https://doi.org/10.37201/REQ/ 071.2021.
- [20] W. Dejnirattisai, R.H. Shaw, P. Supasa, C. Liu, A.S. Stuart, A.J. Pollard, X. Liu, T. Lambe, D. Crook, D.I. Stuart, J. Mongkolsapaya, J.S. Nguyen-Van-Tam, M.D. Snape, G.R. Screaton, Reduced neutralization of SARS-CoV-2 omicron B.11.529 variant by post-immunisation serum, Lancet 399 (2022) 234–236, https://doi.org/10.1016/S0140-6736(21)02844-0.
- [21] Y. Araf, F. Akter, Y. Tang, R. Fatemi, M. Sorwer Alam Parvez, C. Zheng, M. Golzar Hossain, Omicron Variant of SARS-CoV-2: Genomics, Transmissibility, and Responses to Current COVID-19 Vaccines, vol. 94, 2022, pp. 1825–1832, https://doi.org/10.1002/jmv.27588.
- [22] W. Dejnirattisai, R.H. Shaw, P. Supasa, C. Liu, A.S. Stuart, A.J. Pollard, X. Liu, T. Lambe, D. Crook, D.I. Stuart, J. Mongkolsapaya, J.S. Nguyen-Van-Tam, M.D. Snape, G.R. Screaton, Reduced neutralization of SARS-CoV-2 omicron B.1.1.529 variant by post-immunisation serum, Lancet 399 (2022) 234–236, https://doi.org/10.1016/S0140-6736(21)02844-0.
- [23] W. Doster, A. Bachleitner, R. Dunau, M. Hiebl, E. Lüscher, Thermal properties of water in myoglobin crystals and solutions at subzero temperatures, Biophys. J. 50 (1986) 213–219, https://doi.org/10.1016/S0006-3495(86)83455-5.
- [24] A.H. Linden, W.T. Franks, Ü. Akbey, S. Lange, B.J. Van Rossum, H. Oschkinat, Cryogenic temperature effects and resolution upon slow cooling of protein preparations in solid state NMR, J. Biomol. NMR. 51 (2011) 283-292, https:// doi.org/10.1007/s10858-011-9535-z.
- [25] R. Sanjuán, P. Domingo-Calap, Mechanisms of viral mutation, Cell. Mol. Life Sci. 73 (2016) 4433–4448, https://doi.org/ 10.1007/s00018-016-2299-6.
- [26] K.T. Bæk, R. Mehra, K.P. Kepp, Stability and expression of SARS-CoV-2 spike-protein mutations, Mol. Cell Biochem. 17 (2022) 1–12, https://doi.org/10.1007/s11010-022-04588-w.
- [27] A. V Kudriavtsev, A. V Vakhrusheva, V.N. Novoseletsky, M.E. Bozdaganyan, K. V Shaitan, M.P. Kirpichnikov, O.S. Sokolova, Immune escape associated with RBD omicron mutations and SARS-CoV-2 evolution dynamics, Viruses 14 (2022) 1603, https://doi.org/10.3390/v14081603.
 [28] N. Jalali, H.K. Brustad, A. Frigessi, E.A. Macdonald,
- [28] N. Jalali, H.K. Brustad, A. Frigessi, E.A. Macdonald, H. Meijerink, S.L. Feruglio, K.M. Nygård, G. Rø, E.H. Madslien, B. Freiesleben De Blasio, Increased household transmission and immune escape of the SARS-CoV-2 Omicron compared to Delta variants, 2022, p. 5706, https:// doi.org/10.1038/s41467-022-33233-9, 13.
- [29] Q. Li, M. Zhang, Z. Liang, L. Zhang, X. Wu, C. Yang, Y. An, J. Tong, S. Liu, T. Li, Q. Cui, J. Nie, J. Wu, W. Huang, Y. Wang, Antigenicity comparison of SARS-CoV-2 Omicron sublineages with other variants containing multiple mutations in RBD, MedComm 3 (2022) 130, https://doi.org/ 10.1002/mco2.130.
- [30] V. Thakur, R.K. Ratho, OMICRON (B.1.1.529): a new SARS-CoV-2 variant of concern mounting worldwide fear, J. Med. Virol. 94 (2022) 1821–1824, https://doi.org/10.1002/ jmv.27541.
- [31] Z. Du, H. Hong, S. Wang, L. Ma, C. Liu, Y. Bai, D.C. Adam, L. Tian, L. Wang, E.H.Y. Lau, B.J. Cowling, Reproduction number of the omicron variant triples that of the Delta variant, Viruses 14 (2022) 4–8, https://doi.org/10.3390/ v14040821.